



## Synthesis of aspergillide A from a synthetic intermediate of aspergillide B

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### ABSTRACT

The first synthesis of aspergillide A, a cytotoxin produced by a marine-derived fungus, has been achieved from a synthetic intermediate of aspergillide B by using a proline-mediated epimerization of a 2,6-trans-substituted tetrahydropyran-2-acetaldehyde intermediate into the corresponding cis-isomer via a retro-oxy-Michael/oxy-Michael sequence as the key transformation.

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In the course of screening for bioactive substances from marine-derived fungi, Kusumi and co-workers isolated three cytotoxic compounds, aspergillides A–C, from a bromine-modified 1/2PD (potato-dextrose) culture medium of *Aspergillus ostianus* strain 01F313, and proposed their structures to be 14-membered macrolides **I**, **II**, and **III**, respectively, based on spectroscopic analyses including NOESY experiments and the modified Mosher method (Fig. 1).<sup>1</sup> The proposed structure of aspergillide C (**III**) was confirmed by our total synthesis,<sup>2</sup> while the stereochemical assignment of aspergillides A and B was found to be incorrect through the total synthesis of **I** and **II** by Hande and Uenishi.<sup>3</sup> They revealed that the physical and spectral data of their synthetic compound **I** were identical to those reported by Kusumi and co-workers for aspergillide B, and the data of synthetic **II** did not match those reported either for aspergillide A or for aspergillide B. These results enabled them to conclude that the genuine structure of aspergillide B must be **2** (Fig. 2), and the real structure of aspergillide A should be reinvestigated. Ooi and co-workers answered the question regarding the structure of aspergillide A by means of X-ray crystallographic analysis of its *m*-bromobenzoate derivative, which clarified that aspergillide A was the C3 epimer of aspergillide B (compound **1**, Fig. 2) possessing a 3,7-cis relationship as well as 3*R* absolute configuration.<sup>4</sup>

Prompted by the unique molecular architecture of aspergillides A (**1**), B (**2**), and C (**III**), the latter two of which possess a very rare 2,6-trans-substituted tetrahydro- and dihydropyran structural unit, respectively, embedded in a 14-membered macrolide structure,<sup>5</sup> three additional syntheses of **2**<sup>6</sup> and an additional synthesis

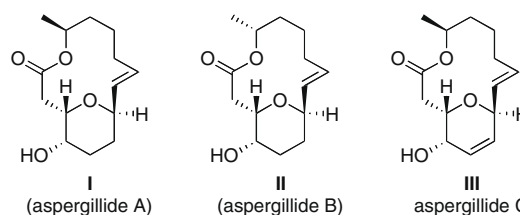


Figure 1. Originally proposed structures of aspergillides A (**I**), B (**II**), and C (**III**).

of **III**<sup>7</sup> have been published. However, no report concerning the synthesis of aspergillide A (**1**) has appeared to date. We describe herein the first synthesis of aspergillide A via a proline-mediated isomerization of a 2,6-trans-substituted tetrahydropyran-2-acetaldehyde intermediate into the corresponding cis-isomer.

Since aspergillide A (**1**) and aspergillide B (**2**) are epimeric to each other at the alkoxy-bearing C3 position  $\beta$  to the lactone carbonyl, we envisaged that they might be interconvertible via a retro-oxy-Michael/oxy-Michael equilibrium sequence. Concerned about the possible formation of a  $\gamma$ -lactone from **2** through the intramolecular attack of the C4 hydroxyl to the carbonyl group

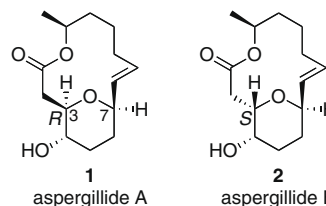
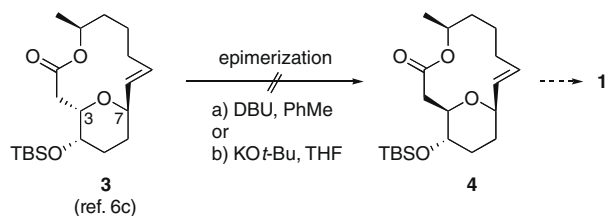


Figure 2. Revised stereochemistries of aspergillides A and B.

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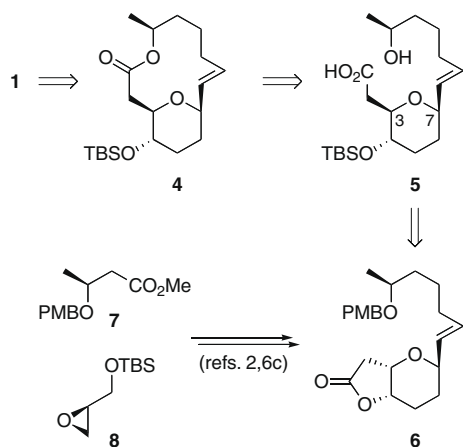


**Scheme 1.** Attempted epimerization of TBS-protected aspergillide B (3) to TBS-protected aspergillide A (4).

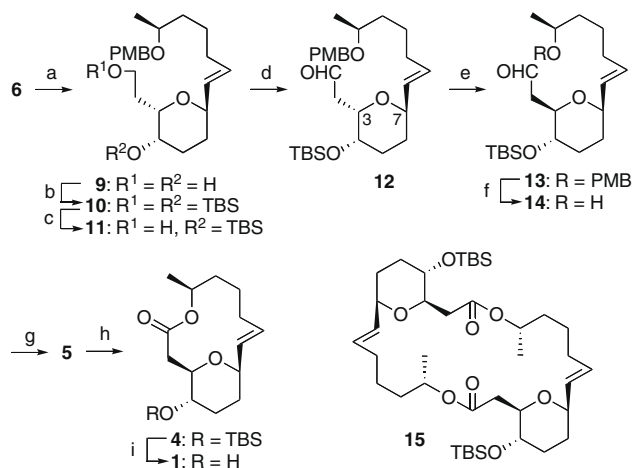
during the attempt to directly epimerize 2 into 1, we chose the TBS-protected form of 2 (compound 3, Scheme 1), the penultimate intermediate in Uenishi's and our syntheses of aspergillide B,<sup>3,6c</sup> as the substrate for the epimerization. Thus, the 3,7-trans isomer 3 was first subjected to DBU in toluene in the hope of obtaining the corresponding 3,7-cis isomer 4, the TBS ether of 1. The basic treatment, however, did not afford the desired product 4 even at elevated temperatures, resulting only in the recovery of the starting material 3. The use of KOt-Bu as a stronger base also brought no fruitful outcome, giving only the starting material at room temperature or a complex mixture at 45 °C.<sup>8</sup>

Faced with the difficulty to epimerize the 3,7-trans macrolactone 3 into the corresponding cis-isomer 4, we next planned to prepare 3,7-cis-substituted seco acid 5 beforehand and then macrolactonize it into 4 (Scheme 2). As a possible precursor of 5, we chose 3,7-trans-substituted lactone 6 since it was known to be obtainable very efficiently from 7 and 8 in our previous synthesis of aspergillide B (2),<sup>2,6c</sup> and the requisite stereochemical inversion at the C3 position of 6 on its way to 5 was expected to be possible from literature precedents by conducting an appropriate epimerization reaction on a suitable intermediate before macrolactonization.<sup>9</sup>

The elaboration of 6 to 1 commenced with the reductive opening of the lactone ring of 6 with LiAlH<sub>4</sub> to give diol 9. Protection of the two hydroxyl groups of 9 to bis-TBS ether 10 was followed by selective removal of the protecting group at the primary hydroxyl, affording 11 in 86% yield for the three steps (Scheme 3). Oxidation of the resulting alcohol with Dess–Martin's periodinane proceeded smoothly to furnish aldehyde 12, which set the stage for the key transformation in the present synthesis, the epimerization at the C3 position of the 3,7-trans-substituted intermediate 12 to the corresponding 3,7-cis isomer 13. The conversion of 12 into 13 was realized very efficiently by the Massi–Dondoni protocol using proline as the epimerization catalyst.<sup>10</sup> Thus, the treatment of 12 with D-proline for 1 h at 0 °C and for an additional 3 h at 60 °C gave a 95:5 epimeric mixture of 13 and 12 in 81% yield favoring the de-



**Scheme 2.** Synthetic plan for aspergillide A (1) from known compound 6.



**Scheme 3.** Conversion of 6 into aspergillide A (1). Reagents and conditions: (a) LiAlH<sub>4</sub>, THF, 0 °C to rt, 1 h; (b) TBSOTf, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C to rt, 1 h; (c) CSA (0.2 equiv), CH<sub>2</sub>Cl<sub>2</sub>/MeOH, 0 °C, 1 h (86%, three steps); (d) Dess–Martin's periodinane, NaHCO<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, rt, 5 h (97%); (e) D-proline (0.3 equiv), MeOH, 0 °C, 1 h, then 60 °C, 3 h (81%); (f) DDQ, phosphate buffer (pH 7.0), CH<sub>2</sub>Cl<sub>2</sub>, rt, 8 h (80%); (g) NaClO<sub>2</sub>, NaH<sub>2</sub>PO<sub>4</sub>, 2-methyl-2-butene, *t*-BuOH/H<sub>2</sub>O, 0 °C, 7 h (97%); (h) Cl<sub>3</sub>C<sub>6</sub>H<sub>2</sub>COCl, Et<sub>3</sub>N, THF, 0 °C to rt, 2 h, then DMAP, PhMe, 80 °C, 8 h (30%); (i) TBAF, THF, rt, 2 h (76%).

sired 3,7-cis isomer 13.<sup>11</sup> After deprotection of the PMB group of 13 by DDQ oxidation, the resulting product 14 was oxidized to the seco acid 5 by the Pinnick oxidation. Unexpectedly, the macrolactonization of 5 into 4 was found to be problematic in contrast to the case of the corresponding 3,7-trans seco acid, which underwent smooth macrolactonization under the Yamaguchi lactonization conditions and led, after deprotection, to aspergillide B (2) in previous synthetic studies.<sup>3,6c</sup> On treatment of 5 with 2-methyl-6-nitrobenzoic anhydride in the presence of DMAP in CH<sub>2</sub>Cl<sub>2</sub> at room temperature for 28 h (Shiina's method),<sup>12</sup> no desired product 4 was obtained, but instead dimeric macrodiolide 15 was isolated in 22% yield. Gerlach's modification of the Corey–Nicolaou macrolactonization (PySSPy, Ph<sub>3</sub>P, AgBF<sub>4</sub>, PhMe, 110 °C, 54 h)<sup>13</sup> and modified Mukaiyama's lactonization conditions (2-bromo-1-ethylpyridinium tetrafluoroborate, Et<sub>3</sub>N, MeCN, 90 °C, 6 h)<sup>14</sup> both gave complex mixtures. The only successful result was obtained when the seco acid 5 was subjected to Yamaguchi's conditions (Cl<sub>3</sub>C<sub>6</sub>H<sub>2</sub>COCl, Et<sub>3</sub>N, THF, then DMAP, PhMe, 81 °C, 8 h; substrate concentration, 0.8 mM),<sup>15</sup> which gave the desired product 4 in 30% yield along with 19% of the dimeric product 15.<sup>16,17</sup> Unfortunately, all attempts to improve the chemical yield of 4 by varying the reaction conditions (mainly, reaction temperature, and concentration) were unsuccessful and could not exceed the above-mentioned yield (30%). Finally, the TBS-protecting group of 4 was removed with TBAF to give aspergillide A (1) as a crystalline solid (mp 64.5–65.5 °C) after chromatographic purification. The specific rotation and spectral data of 1 were in good agreement with those reported in the literature.<sup>1,18</sup>

In conclusion, the first synthesis of aspergillide A (1) was accomplished from 6, an intermediate in our total synthesis of aspergillide B, by using the proline-mediated epimerization of the 3,7-trans-substituted cyclic intermediate 12 into the corresponding cis-isomer 13 as the key step. Efforts to develop a more efficient synthetic route to 1, including the improvement of the macrolactonization step, are now underway and will be reported in due course.

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